


  
**GENOMICS EDUCATION PARTNERSHIP**

## Long-Read Sequencing Technology

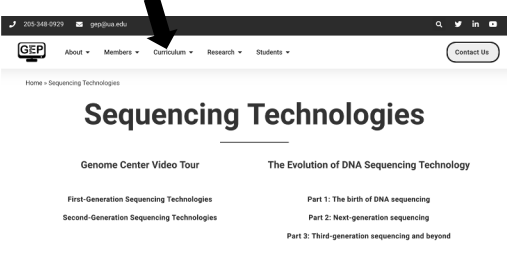
Katie M. Sandlin, M.S.

 [kmsandlin@ua.edu](mailto:kmsandlin@ua.edu)  
 @KatieMSandlin  
 [linkedin.com/in/kmsandlin](https://www.linkedin.com/in/kmsandlin)


June 20, 2024

1

## Currently Available GEP Resources




[thegep.org/sequencing/](https://thegep.org/sequencing/)



2

## Note to Instructors


- I have scripted the talk in the notes section of the PPT.
- Advanced versions of this presentation and lecture recordings were developed by Wilson Leung (Washington University in St. Louis).
  - [Overview Third-generation Sequencing](#)
    - [Lecture Recording](#)
  - [Overview PacBio Iso-Seq](#)
    - [Lecture Recording](#)



3

## Agenda

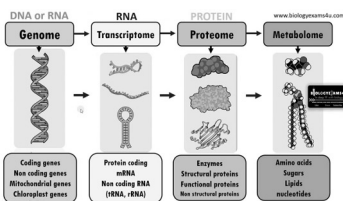
1. Review of RNA-Seq Short-read Sequencing
2. Overview and benefits of Long-read Sequencing
3. Oxford Nanopore Sequencing
4. Pacific Biosciences Sequencing



4


## RNA Sequencing (RNA-Seq)

- RNA sequencing (or RNA-seq) lets us investigate and discover the transcriptome—the total cellular content of RNAs including mRNA, rRNA and tRNA—by using next-generation sequencing to examine the quantity and sequences of RNA in a sample.
  - RNA-seq analyzes the transcriptome, indicating which of the genes encoded in our DNA are turned on or off and to what extent.



The **transcriptome** is the complete set of transcripts in a cell, and their quantity, for a specific developmental stage or physiological condition.


Wang, Z., Gerstein, M., & Snyder, M. (2009). <https://doi.org/10.1016/j.cell.2009.02.004>




5

## RNA-Seq: reads are typically 30–300 bp

① Isolate RNA from samples





6

**Transcriptome reconstruction—akin to reassembling magazine articles after they have been through a paper shredder**

Goodell, J. (2012). Genomics: the state of the art in RNA-seq analysis. *Nature Methods*. 7

7

**Short vs. Long Read RNA-Seq**

Adapted from: *Genetic Engineering & Biotechnology News*. 8

8

**Agenda**

1. Review of RNA-Seq Short-read Sequencing
2. Overview and benefits of Long-read Sequencing
3. Oxford Nanopore Sequencing
4. Pacific Biosciences Sequencing

Pacific Biosciences 9

9

**Short vs. Long Read Sequencing**

Sequencing instruments are categorized as long-read or short-read based on their underlying chemistry and the length of DNA fragments they analyze.

PacBio HiFi Sequencing - Unlock Your Next Great Discovery 10

10

**Benefits of assembling a genome with long-read sequencing: an example using books**

- A way to think about the difference between assembling a genome with short-reads versus long-read sequencing technology would be to imagine two different approaches to reconstructing a 368-page novel from randomized snippets of text.

Pacific Biosciences 11

11

**Benefits of assembling a genome with long-read sequencing: an example using books**

Those looking for serious land

the North Carolina coast

The shack sat back

hatchet and pack a buck

by a torn shoreline

Excerpts from: *Where the Crawdads Sing* 12

12

### Benefits of assembling a genome with long-read sequencing: an example using books

The marsh was guarded by a twin shoalway, labeled by early explorers as the "Crusoyard of the Atlantic" because rippling, furious winds, and shallow shoals wrecked ships like paper bats along what would become the North Carolina coast. One seaman's journal read, "rang'd along the Shoar . . . but could discover no Entrance . . . A violent Storm overtook us . . . we were forced to get off to Sea, to secure Chives and Ship, and were driven by the Rapidity of a strong Current . . ."

*Where the Crawdads Sing*

THE SHACK SAT BACK from the palmettos, which sprawled across sand flats to a necklace of green lagoons and, in the distance, all the marsh beyond. Miles of blade grass so tough it grew in salt water interrupted only by trees as best they were the shape of the wind. Oak forests branched around the other sides of the shack and sheltered the closest lagoons; its surface so rich in life it churned. Salt air and gull-song drifted through the trees from the sea.

"The Land . . . being marshy and Swamps, we return'd towards our Ship . . . Discouragement of all such as should hereafter come into these Parts to settle."

Claiming territory hadn't changed much since the 1600s. The scattered marsh holdings weren't legally described, just marked out natural—a creek boundary here, a dead oak there—by megalodes. A man doesn't set up a palmetto lean-to in a bog unless he's on the run from somebody or at the end of his own road.

Those looking for arroyo land moved on, and that infamous marsh became a nest, scooping up a mishmash of mutinous sailors, castaways, debtors, and fugitives dodging wars, taxes, or laws that they didn't like to. The ones malaria didn't kill or the swampy didn't swallow bred into a woodsmen tribe of several races and multiple cultures, each of whom could fill a small frame with a hatchet and pack a buck for miles. Like

Excerpts from *Where the Crawdads Sing* 13

13

### Benefits of assembling a genome with long-read sequencing: an example using books

The shack sat back from the palmettos, which sprawled across sand flats to a necklace of green lagoons and, in the distance, all the marsh beyond. Miles of blade grass so tough it grew in salt water interrupted only by trees as best they were the shape of the wind. Oak forests branched around the other sides of the shack and sheltered the closest lagoons; its surface so rich in life it churned. Salt air and gull-song drifted through the trees from the sea.

*Where the Crawdads Sing*

THE SHACK SAT BACK from the palmettos, which sprawled across sand flats to a necklace of green lagoons and, in the distance, all the marsh beyond. Miles of blade grass so tough it grew in salt water interrupted only by trees as best they were the shape of the wind. Oak forests branched around the other sides of the shack and sheltered the closest lagoons; its surface so rich in life it churned. Salt air and gull-song drifted through the trees from the sea.

"The Land . . . being marshy and Swamps, we return'd towards our Ship . . . Discouragement of all such as should hereafter come into these Parts to settle."

Claiming territory hadn't changed much since the 1600s. The scattered marsh holdings weren't legally described, just marked out natural—a creek boundary here, a dead oak there—by megalodes. A man doesn't set up a palmetto lean-to in a bog unless he's on the run from somebody or at the end of his own road.

Those looking for arroyo land moved on, and that infamous marsh became a nest, scooping up a mishmash of mutinous sailors, castaways, debtors, and fugitives dodging wars, taxes, or laws that they didn't like to. The ones malaria didn't kill or the swampy didn't swallow bred into a woodsmen tribe of several races and multiple cultures, each of whom could fill a small frame with a hatchet and pack a buck for miles. Like

Excerpts from *Where the Crawdads Sing* 14

14

### 3rd Generation (Long-Read) Sequencing Emerges

- To address the limitations of next-generation sequencing, scientists began developing real-time, single-molecule DNA sequencing platforms.
- In the mid 2000s, two single-molecule technologies emerged giving rise to 3rd generation sequencing:
  - single-molecule real-time (SMRT) sequencing by Pacific Biosciences
  - nanopore sequencing by Oxford Nanopore Technologies

First generation: Sanger sequencing (Maxam and Gilbert, Bigelow DNA sequencing). Second generation: High throughput from the parallelization of sequencing reactions (454, Solexa, Ion Torrent, Illumina). Third generation: Sequence native DNA in real-time with single-molecule resolution (Pacific Oxford Nanopore).

Short-read sequencing vs Long-read sequencing

15 Pacific Biosciences

15

### Third-generation Sequencing Technologies

- Sequencing platforms from Pacific Biosciences (PacBio) and Oxford Nanopore Technologies (ONT)
- Key characteristics:
  - Single-molecule, real-time sequencing
    - Sequence samples directly without amplification
  - Can detect epigenetic modifications
    - DNA methylation
- Comparisons with Illumina:
  - Produce **longer reads**
  - Raw reads have **higher error rates** than Illumina

16

16

## Agenda

1. Review of RNA-Seq Short-read Sequencing
2. Overview and benefits of Long-read Sequencing
3. Oxford Nanopore Sequencing
4. Pacific Biosciences Sequencing

17

17

### ONT Sequencing – Template Topology

- Yellow – forward DNA strand
- Dark Blue – reverse DNA strand
- Light Blue – sequencing adapters

Adapter-tagged DNA, 1 kb to >2 Mb fragment, Motor protein

18

18

## ONT Sequencing – Flow Cell

- Yellow – forward DNA strand
- Dark Blue – reverse DNA strand
- Light Blue – sequencing adapters

Flow cell (top view)

Nanopore

Synthetic membrane

Logsdon, G.A., Volgger, M.R. & Eichler, E.E. [Long-read human genome sequencing and its applications. Nat Rev Genet 21, 597–614 \(2020\).](#) 19

19

## ONT Sequencing - Nanopore

- Yellow – forward DNA strand
- Dark Blue – reverse DNA strand
- Light Blue – sequencing adapters

Single nanopore (cross section)

Motor protein

Nanopore

Voltage-biased membrane

Logsdon, G.A., Volgger, M.R. & Eichler, E.E. [Long-read human genome sequencing and its applications. Nat Rev Genet 21, 597–614 \(2020\).](#) 20

20

## ONT Sequencing – Base Calling

- You can think of sequencing DNA or RNA like reading music — the bases being the notes, which when played in the correct order, produces a recognizable song.

Readout

Mean signal (pA)

Time (seconds)

Logsdon, G.A., Volgger, M.R. & Eichler, E.E. [Long-read human genome sequencing and its applications. Nat Rev Genet 21, 597–614 \(2020\).](#) 21

21

## Check For Understanding

Raise your fingers (1-4) to share your answer.

1. Synthetic Membrane
2. Motor Protein
3. DNA or RNA
4. Nanopore

Cold Spring Harbor Laboratory – DNA Learning Center. 22

22

## Check For Understanding

Raise your fingers (1-4) to share your answer.

1. Synthetic Membrane
2. Motor Protein
3. DNA or RNA
4. Nanopore

Cold Spring Harbor Laboratory – DNA Learning Center. 23

23

## Check For Understanding

Raise your fingers (1-4) to share your answer.

1. Synthetic Membrane
2. Motor Protein
3. DNA or RNA
4. Nanopore

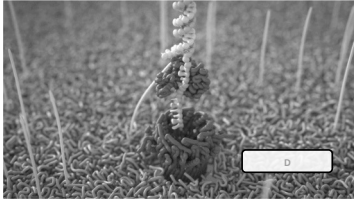
Cold Spring Harbor Laboratory – DNA Learning Center. 24

24

## Check For Understanding

Raise your fingers (1-4) to share your answer.

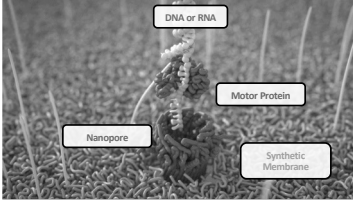
1. Synthetic Membrane
2. Motor Protein
3. DNA or RNA
4. Nanopore



Cold Spring Harbor Laboratory – DNA Learning Center

25

## Check For Understanding



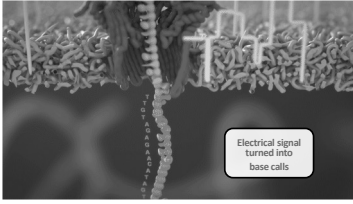
- Read length is up to 4 Mb
  - Long reads sequencing mode: 10–100kb
  - Ultra-long sequencing mode: 100–300kb

Cold Spring Harbor Laboratory – DNA Learning Center

26

## Review – Base Calling

- This digital method of reading DNA or RNA sequence has multiple benefits:
  - Real-time analysis
  - PCR free, no amplification bias
  - **Modified base detection**
  - Read-length agnostic
  - **Direct sequencing of DNA or RNA**




Disclaimer: The real-time base caller has lower accuracy. For many applications, it might be beneficial to do the base calling afterwards with a more powerful GPU and a more compute intensive algorithm [i.e., duplex with super accuracy (SUP)].

Cold Spring Harbor Laboratory – DNA Learning Center


27

## Flow Cell


### Where Nanopores are Housed



The **Flongle** Flow Cell can generate up to 2.8 Gb of data enabling direct, real-time DNA & cDNA sequencing on smaller, single-use flow cells.



The **MinION** Flow Cell can generate up to 50 Gb of data for sequencing DNA, cDNA or native RNA in real-time.

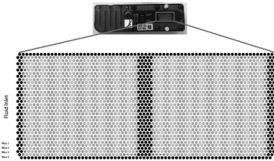


The **PromethION** Flow Cell can generate up to 290 Gb for sequencing DNA, cDNA or native RNA in real-time.

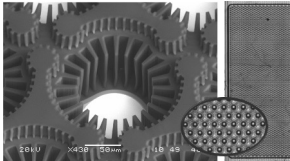
https://nanoporetech.com/platform/technology/flow-cells-and-nanopores

28

## MinION Flow Cell



MinION Mk1B and Mk1C/GridION Flow Cell Channel Layout




SEM image of a single sensor well (left) and the hexagonal array of wells (right).

Oxford Nanopore Technologies

29

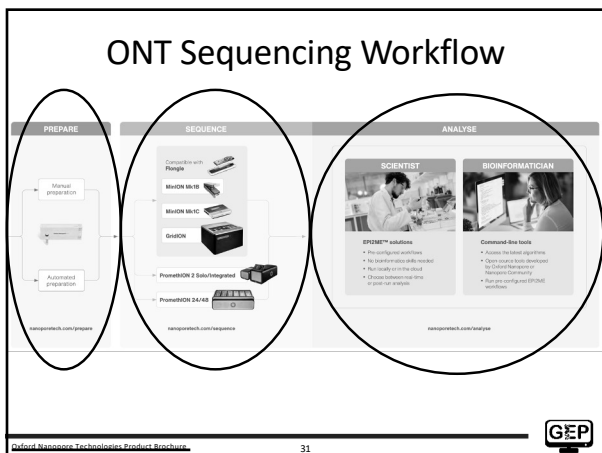
## Nanopore Sequencing Devices



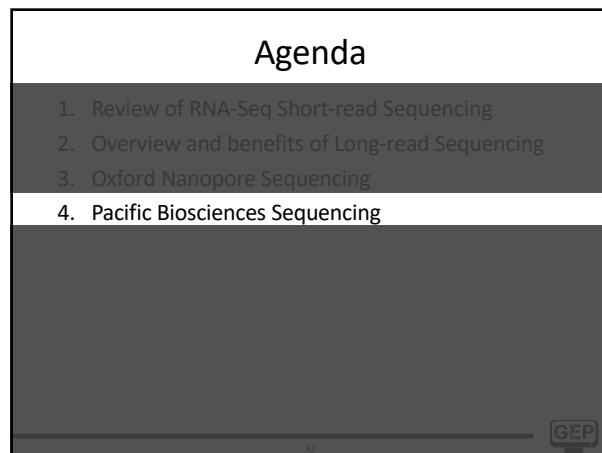
**\$1,999**

Oxford Nanopore Technologies

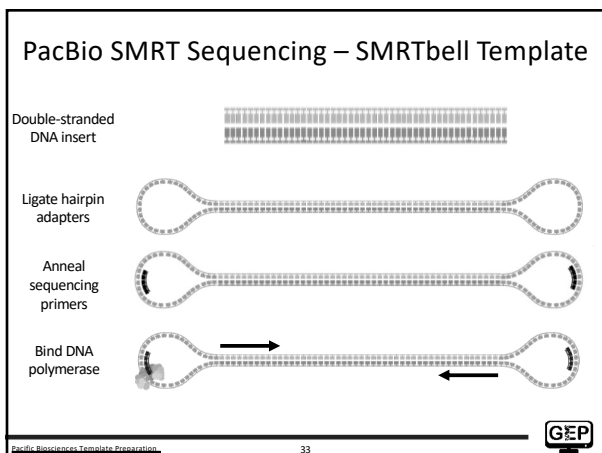
30



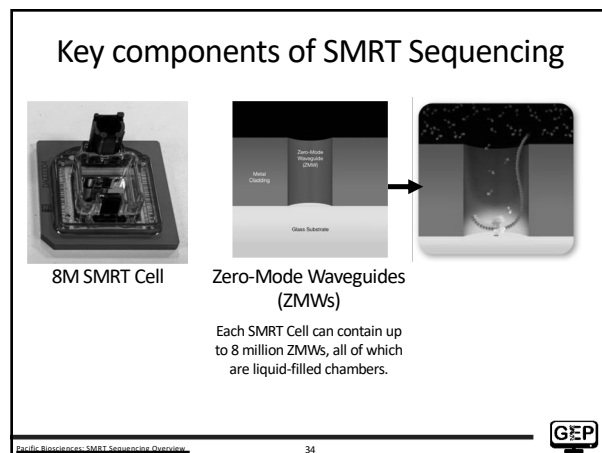
31



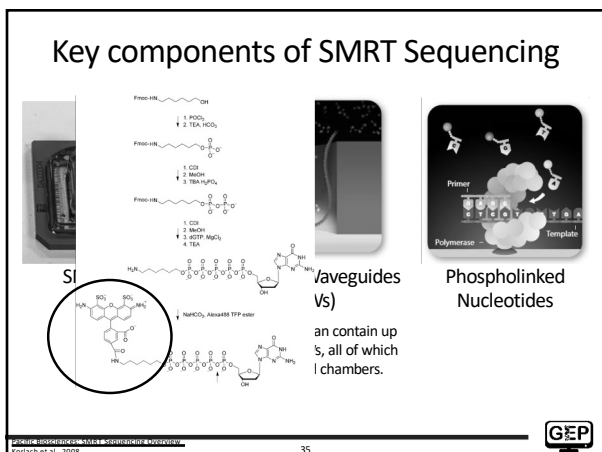
32



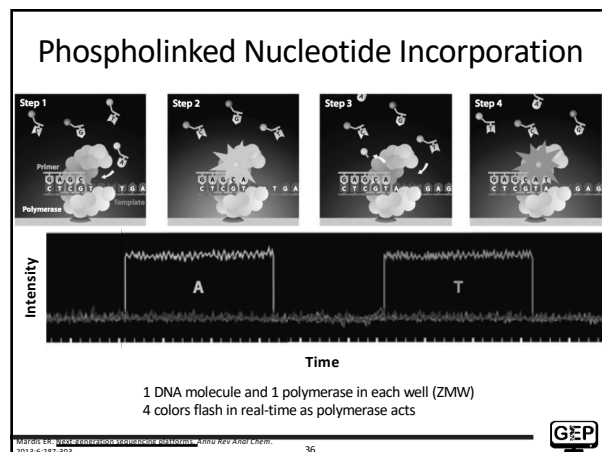
33



34



35

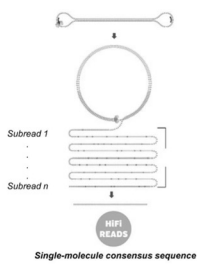


36

## 2 Modes of SMRT Sequencing

### Circular Consensus Sequencing (CCS) Mode

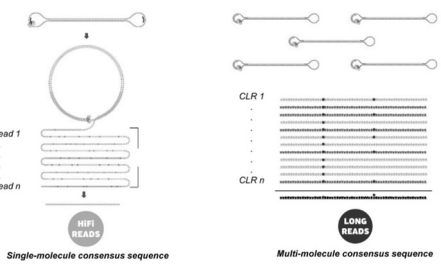
Inserts 10-20 kb



Single-molecule consensus sequence

### Continuous Long Read (CLR) Sequencing Mode

Inserts >25 kb, up to 175 kb



Multi-molecule consensus sequence

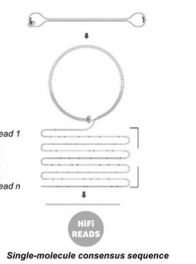
Pacific Biosciences

37

## 2 Modes of SMRT Sequencing

### Circular Consensus Sequencing (CCS) Mode

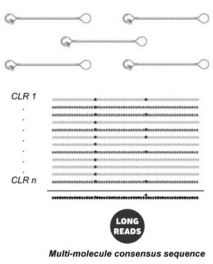
Inserts 10-20 kb



Single-molecule consensus sequence

### Continuous Long Read (CLR) Sequencing Mode

Inserts >25 kb, up to 175 kb



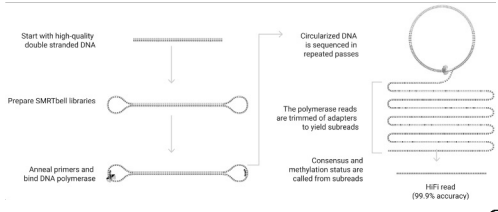
Multi-molecule consensus sequence

Pacific Biosciences

38

## How are HiFi reads generated?

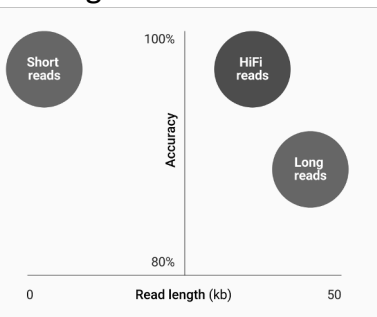
- SMRT sequencing can sequence the same DNA molecule **multiple times** generating highly accurate long reads, or **HiFi reads**.
- HiFi reads are produced using **circular consensus sequencing (CCS)** mode on PacBio long-read systems. HiFi reads provide base-level resolution with 99.9% single-molecule read accuracy.



Pacific Biosciences – HiFi Sequencing

39

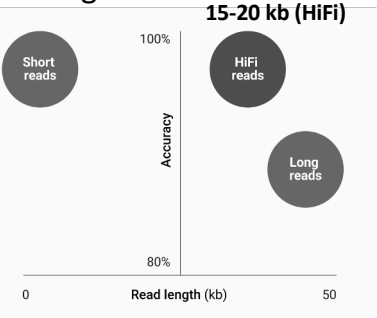
## HiFi Reads vs. Long Reads



Pacific Biosciences – HiFi Sequencing

40

## HiFi Reads vs. Long Reads



**15-20 kb (HiFi)**

**Disclaimer:** PacBio HiFi reads typically have lower quality than Illumina data. They also have systematic errors (particularly with homopolymers and specific types of repetitive sequences; see [McFarmer et al., 2022](#)).


PacBio HiFi reads are defined as reads with an accuracy of Q20 or above (error rate of 1/100; 99% accuracy). For the Revio system, the mode for the read quality is ~Q30 (99.9% accuracy; see the [Revio system brochure](#)).

For the Illumina XLEAP-SBS chemistry, the read quality is Q40 (99.99% accuracy; see the [“XLEAP-SBS chemistry enables Q40 and above data quality on NovaSeq X and NextSeq 1000/2000”](#) blog post).

Pacific Biosciences – HiFi Sequencing

41

## PacBio Sequencing – How it Works



©PacBio PacBio Sequencing – How it Works

42

### Overview of PacBio Sequencers

**Sequel**

**Revio One Work Deck**

- Pipettor
- Trap door
- Tri-position sensor
- Auxiliary tip holder
- Sequencing plate reagent slots 1 and 2
- Test cell and leak test pad
- Robot
- Cell prep station
- Evaporation lid
- SMRT Cell tray slot
- Tip box slot
- Waste bin
- Work deck touchscreen

**1M ZMW**

**8M ZMW**

**25M ZMW**

**SMRT Cell tray**

Images provided by Pacific Biosciences

43

### PacBio Revio Sequencing System

**\$779,000**

Well Connected

44

### PacBio SMRT Sequencing Workflow

Although each sequencing project is unique, there are three main steps to go from DNA to discovery with SMRT sequencing.

- Library prep**  
[Learn more about kits for fast and easy library preparation](#)
- SMRT sequencing**  
[Explore sequencing interactive](#)
- Data analysis**  
[SMRT Link user interface and SMRT Analysis](#)

Pacific Biosciences - Steps of SMRT Sequencing

45

### Isoform Sequencing (Iso-Seq)

- Using SMRT sequencing to do RNA-Sequencing = marketing term is Iso-Seq
- Iso-Seq data is obtained via Circular Consensus Sequencing (CCS)

**Iso-Seq**

Scalable  
De Novo  
Isoform Discovery  
from PacBio HiFi Reads

46

### Benefits of Long-read RNA-Seq

Gene n

Transcript x

Transcript y

**Illumina RNA-Seq**

- Short-read cDNA
- Ambiguous to exon
- Unambiguous to exon
- Ambiguous to isoform
- Unambiguous to isoform

**PacBio Iso-Seq**

- Long-read cDNA
- Unambiguous to isoform

**Nanopore**

- Direct RNA-Seq
- Unambiguous to isoform

Reads that map to exons    Reads that map across a splice junction

m<sup>6</sup>A = N<sup>6</sup>-methyladenosine

Stark et al., Nature Reviews Genetics, 2021(1), 431-456

47

### Projects typically combine multiple sequencing technologies to construct genome assemblies

- The Vertebrate Genomes Project (VGP) aims to construct reference genome assemblies for ~70,000 vertebrate species.
- Sequencing technologies used by the Vertebrate Genome Lab (VGL) at the Rockefeller University:

- PacBio Sequel IIe**: Use HiFi data to construct the initial assembly
- ONT Promethion**: Use ultra-long Nanopore reads to resolve highly repetitive regions
- Bionano Saphyr**: Use optical maps to determine order and orientation of the contigs (scaffolding)
- Arima Hi-C**: Second round of scaffolding to generate chromosome-scale assemblies

- Illumina data also used to polish the consensus sequence

VGL Technologies


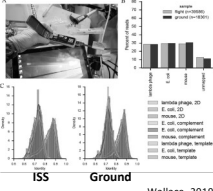
48



### Applications of 3<sup>rd</sup> Generation Sequencing

- Chromosome-scale genome assemblies
- Cancer genomics, human disease research
- Population genetics (SNVs, phasing)
- Epigenomics (DNA methylation)
- Full-length RNA-Seq
  - PacBio Iso-Seq, Nanopore direct RNA-Seq
- Single-cell RNA-Seq
  - PacBio Kinnex (MAS-Seq)
- Conservation genomics
- Complete plasmid verification
  - Plasmidsaurus, Eurofins, ...
- Metagenomics, microbial genomics
- ... ..

**Astronaut Kate Rubins on the ISS**

Wallace, 2019

49

### Conclusion

- Sequencing technology is the ‘microscope’ by which geneticists study genetic variation, and it is clear that long-read technologies have provided us with a new ‘lens and objective’ for understanding DNA and RNA variation, structure and organization.
- Although the two predominant long-read technologies are competitive, some of the best results have been obtained when the sequencing platforms are used to complement one another.

50

### Copy Google Doc Worksheet

	Oxford Nanopore Technologies (ONT)	Pacific Biosciences (PacBio)
Sequencing chemistry/technology name		
Read length		
What is detected in the sequencing process?		
How is that detected?		
Enzyme used in the sequencing process		

[thegep.org/lrs](https://thegep.org/lrs)

51

### Answer Key

	Oxford Nanopore Technologies (ONT)	Pacific Biosciences (PacBio)
Sequencing chemistry/technology name	Nanopore Sequencing	Single Molecule, Real-Time (SMRT)
Read length	Up to 4 Mb; Long reads sequencing mode: 10–100kb; Ultra-long sequencing mode: 100–300kb	15-20 kb (HiFi)
What is detected in the sequencing process?	Changes in current as nucleotides pass through the pore	Fluorophore associated with the phospholinked nucleotides emit different color light
How is that detected?	Different collection of nucleotides that occupy a nanopore will cause different changes in the electric current. Machine learning algorithms (neural networks) are used to analyze the continuous changes to the electric current (squiggle) to infer the sequences that pass through the nanopore.	Each phospholinked nucleotide (A, C, G, T) is labeled with a different fluorescent dye. When a nucleotide is incorporated by the polymerase, it emits different color light that corresponds to the fluorescent dye for the nucleotide.
Enzyme used in the sequencing process	Motor protein (helicase)	DNA polymerase


52

### Acknowledgements

- Special thanks to Wilson Leung (Washington University in St. Louis) for his suggestions and feedback.
- Thank you to Jason Williams and Anna Feitzinger at the CSHL DNA Learning Center for leading a Nanopore Sequencing Workshop.
- This presentation is based upon work supported by the National Science Foundation under Grant No. 1915544 and National Institute of General Medical Sciences of the National Institutes of Health under award number R25GM130517 to LKR.

53

### Questions?



54